



Review

Targeting the Linear Ubiquitin Assembly Complex to Modulate the Host Response and Improve Influenza A Virus Induced Lung Injury[☆]

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ABSTRACT

Influenza virus infection is characterized by symptoms ranging from mild congestion and body aches to severe pulmonary edema and respiratory failure. While the majority of those exposed have minor symptoms and recover with little morbidity, an estimated 500,000 people succumb to IAV-related complications each year worldwide. In these severe cases, an exaggerated inflammatory response, known as “cytokine storm”, occurs which results in damage to the respiratory epithelial barrier and development of acute respiratory distress syndrome (ARDS). Data from retrospective human studies as well as experimental animal models of influenza virus infection highlight the fine line between an excessive and an inadequate immune response, where the host response must balance viral clearance with exuberant inflammation. Current pharmacological modulators of inflammation, including corticosteroids and statins, have not been successful in improving outcomes during influenza virus infection. We have reported that the amplitude of the inflammatory response is regulated by Linear Ubiquitin Assembly Complex (LUBAC) activity and that dampening of LUBAC activity is protective during severe influenza virus infection. Therapeutic modulation of LUBAC activity may be crucial to improve outcomes during severe influenza virus infection, as it functions as a molecular rheostat of the host response. Here we review the evidence for modulating inflammation to ameliorate influenza virus infection-induced lung injury, data on current anti-inflammatory strategies, and potential new avenues to target viral inflammation and improve outcomes.

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Tratamiento dirigido al complejo de ensamblaje de cadenas lineales de ubiquitina para modular la respuesta del huésped y mejorar el daño pulmonar inducido por el virus de la gripe A

RESUMEN

La infección por el virus de la gripe se caracteriza por síntomas que van desde la congestión leve y los dolores corporales hasta el edema pulmonar grave y la insuficiencia respiratoria. Aunque que la mayoría de las personas expuestas presentan síntomas leves y se recuperan con poca morbilidad, se estima que cada año 500.000 personas en todo el mundo fallecen por las complicaciones relacionadas con esta infección. En estos casos graves, se produce una respuesta inflamatoria exagerada, conocida como «tormenta de citocinas», que causa daños en la barrera epitelial respiratoria y el desarrollo del síndrome de distrés respiratorio agudo. Los datos de estudios retrospectivos en humanos, así como de modelos animales experimentales de infección por el virus de la gripe, resaltan la delgada línea que existe entre una respuesta inmunitaria excesiva y una inadecuada, cuando la respuesta del huésped debe mantener el equilibrio entre el aclaramiento viral y la inflamación exagerada. Los moduladores farmacológicos de la inflamación actuales, incluidos los corticoides y las estatinas, no han tenido éxito a la hora de mejorar

Palabras clave:

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los resultados de la infección por el virus de la gripe. Hemos publicado que la amplitud de la respuesta inflamatoria está regulada por la actividad del complejo de ensamblaje de cadenas lineales de ubiquitina (LUBAC, por sus siglas en inglés) y que la atenuación de la actividad de LUBAC protege durante la infección grave por este virus. La modulación terapéutica de la actividad de LUBAC puede ser crucial para mejorar los resultados, ya que funciona como un reóstato molecular de la respuesta del huésped. Aquí revisamos la evidencia al respecto de la modulación de la inflamación para mejorar el daño pulmonar inducido por la infección por el virus de la gripe, los datos sobre las estrategias antiinflamatorias actuales y las posibles nuevas vías para tratar la inflamación viral y mejorar los resultados.

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Introduction

Seasonal influenza A viral infection affects a significant proportion of the population worldwide, with an estimated 500,000 people succumbing to IAV-related complications each year. While most patients infected with influenza A virus (IAV) recover without major sequelae, severe viral pneumonia is one of the most common causes of acute respiratory distress syndrome (ARDS).^{1–4} Clinically, ARDS presents with bilateral pulmonary infiltrates, hypoxemia, pulmonary edema and widespread lung inflammation that lead to high mortality rates due respiratory and to multiple organ failure.^{1,4–6} ARDS patients can be sub-grouped based on the severity of the inflammatory response, where patients with hyper-inflammation have worse clinical outcomes, spending more days on mechanical ventilation, experiencing increased incidence of organ failure and a higher mortality rate compared to hypo-inflammatory ARDS patients.⁷ Impairment of gas exchange in IAV-induced ARDS, in large part, is due to damage to the respiratory epithelial barrier and edema accumulation.^{1,4–8} During IAV infection, an exaggerated inflammatory response, known as “cytokine storm”, can occur leading to the development of hyper-inflammatory ARDS, increasing IAV-induced morbidity and mortality (Fig. 1).⁴ Post-mortem studies of lungs from IAV-infected patients show extensive diffuse alveolar damage characterized by edema, cellular infiltration, thickening of alveolar walls, and necrosis.⁹ Interestingly, a study of critically ill patients showed no differences in pulmonary viral load between those who died and those who recovered, while mortality directly correlated with exuberant inflammation, further supporting a maladaptive host response as the major driver of IAV-induced lung injury.^{10–14} Similar observations are being reported in patients with severe coronavirus disease (COVID-19), where severe lung damage is associated with increased pro-inflammatory cytokines and respiratory failure from ARDS is the leading cause of mortality.^{15,16} With no virus-specific treatment options currently validated, therapies

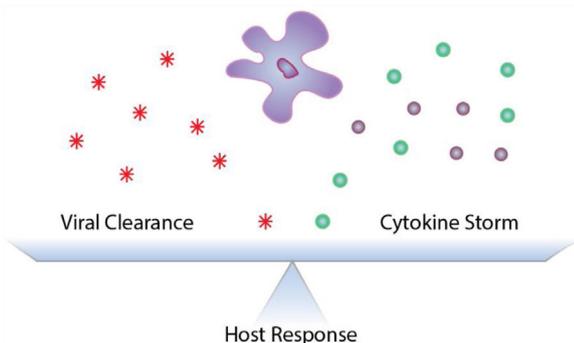


Fig. 1. Schematic representation of the balance that must exist to fine tune the pulmonary inflammatory response to viral infection to promote pathogen clearance and prevent immunopathology.

which target the inflammatory response are currently being considered for patients with severe COVID-19.^{17,18} However, as it has been reported for severe IAV infections, current anti-inflammatory drugs have pleiotropic effects and may lack the specificity needed to carefully calibrate the host response.

Respiratory epithelial cells, as primary targets for IAV infection and replication, initiate inflammatory signaling.^{19–22} In response to respiratory epithelial derived cytokines, innate immune cells, such as neutrophils, monocyte-derived inflammatory macrophages, and natural killer (NK) cells are recruited to the airspace.²³ Together with tissue resident alveolar macrophages, recruited innate immune cells are critical for control of viral replication through both lysis and clearance of virus-infected cells.^{23–26} However, in addition to controlling viral spread, innate immune cells contribute to the overproduction of pro-inflammatory cytokines that enhance IAV-induced lung injury (Fig. 2). The pulmonary immune response must be carefully balanced, simultaneously promoting viral clearance and limiting excessive inflammation to maintain proper lung function.

Findings from animal models of IAV infection have shown that modulation of the host immune response is associated with reduced lung injury and improved survival.^{11,14,27,28} Blockade of

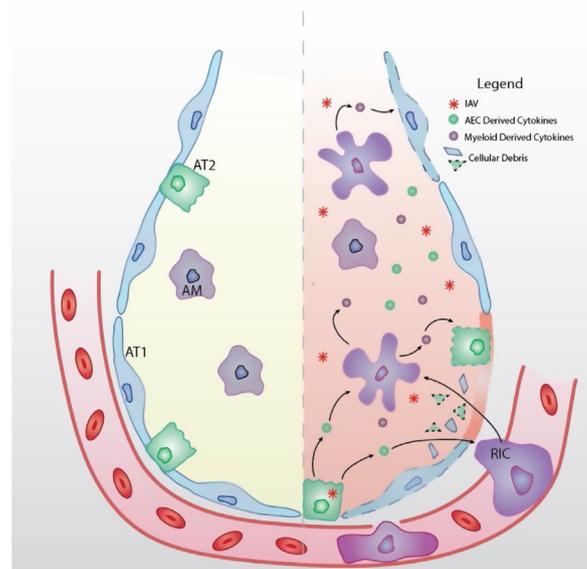


Fig. 2. Schematic model of IAV-induced ARDS (Left) The healthy alveolus is free of fluid with tight junctions between alveolar type 1 (AT1) and type 2 (AT2) epithelial cells. Tissue resident alveolar macrophages (AM) facilitate lung homeostasis through clearance of cellular and environmental debris. (Right) Upon inhalation, IAV targets alveolar epithelial cells (AEC) for infection and replication. IAV infection induces epithelial cytokine production that recruits immune cells to the airspace. Recruited immune cells (RIC) contribute to hyper-inflammatory environment and which further contributes to cytokine storm and tissue damage (dashed outline). Barrier disruption due to the denuded epithelium allows extravasation of proteinaceous edema fluid into the airspace that disrupts gas exchange and causes ARDS.

specific immune cell subsets has been shown to improve outcomes in mouse models of severe IAV infection. For example, genetic deletion of the chemokine receptor CCR2 inhibited the recruitment of monocyte-derived inflammatory macrophages during IAV infection and resulted in reduced lung injury with improved survival. However, loss of this myeloid cell population resulted in a delay in viral clearance.^{29,30} Moreover, adoptive transfer of NK cells from IAV-infected lungs, as compared to NK cell from naïve lungs, resulted in increased mortality of influenza-infected mice,³¹ suggesting that inflammation-dependent activation, rather than recruitment, drives the observed pathology. Taken together data from these animal models suggest that the determinant of influenza severity be orchestrated by respiratory epithelial cells.

LUBAC regulates the amplitude of the lung epithelial driven responses during IAV infection

The linear ubiquitin assembly complex (LUBAC) is a multi-protein E3 ubiquitin ligase complex composed of two stabilizing proteins, the Heme-Oxidized Iron responsive element binding protein 2 ubiquitin Ligase-1L (HOIL-1L) and Shank-Associated RH domain-Interacting Protein (SHARPIN), and a catalytic component, HOIL-1-Interacting Protein (HOIP)^{32–34} (Fig. 3). The proteins within the heteromeric complex contain multiple domains for interactions within the complex, ubiquitin binding, as well as catalytic activity.^{32–36} LUBAC is an essential regulator of NF- κ B activation and has been shown to act as a molecular rheostat, regulating the amplitude of the epithelial-driven inflammatory response during IAV infection.^{35,36}

The respiratory epithelium actively participates in the first line of defense against pathogens by orchestrating host innate immunity.^{23,37–39} As IAV replicates within the respiratory epithelium cells, the cytosolic pattern recognition receptor (PRR), RIG-I, is activated and initiates formation of a signaling platform to which LUBAC is recruited. LUBAC covalently attaches Met-1 linked linear ubiquitin chains to the NF- κ B essential modulator (NEMO), a component of the inhibitor of NF- κ B (I κ B) kinase (IKK) complex along with IKK α and IKK β .^{40,41} Due to the high affinity of NEMO's ubiquitin binding domain for linear chains, linear ubiquitination of NEMO facilitates the recruitment of additional IKK complexes, which results in the efficient trans-autophosphorylation and activation of proximal IKK β followed by the phosphorylation and degradation of I κ B α and robust NF- κ B activation (Fig. 3).^{34,35,40,42,43}

Recent reports show that destabilization of respiratory epithelial LUBAC, via loss of the non-catalytic component HOIL-1L, dampens the host response during severe influenza and promotes survival with reduced lung injury as well as reduced viral titers. However, when LUBAC activity is abolished through deletion of HOIP, the alveolar epithelia driven inflammatory response is inhibited and mortality is increased. These findings highlight the fine line between an excessive and an inadequate immune response and suggest that therapeutic modulation of LUBAC activity may be crucial, as it functions as a rheostat regulating the amplitude of the host response to IAV infection.

LUBAC covalently attaches linear ubiquitin chains to NEMO, which facilitates the recruitment of additional IKK complexes. Stably docked IKK complexes result in the efficient transautophosphorylation and activation of proximal IKK α/β , followed by the phosphorylation and degradation of I κ B α . NF- κ B translocates to the nucleus to stimulate transcription of inflammatory genes.

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Pharmacological immunomodulation during IAV infection

Current anti-influenza strategies are limited to yearly vaccination or administration of antiviral drugs, however, short therapeutic windows, viral mutation, and resistance to current therapies limit their effectiveness. Despite available vaccination and anti-viral drugs, the most recent pandemic in 2009 resulted in an estimated 151,700–575,400 deaths in its first year of circulation worldwide.⁴⁴ The pandemic strain contained a novel assortment of viral genes not previously identified in animal or human populations. From its first detection in April 2009, it was only 3 months until resistance to anti-viral drugs was reported, and it took an additional 3 months before the first vaccine offering protection from the pandemic strain was administered.⁴⁵ In addition to emerging pandemic strains, between 291,000 and 646,000 people worldwide die from seasonal influenza-related respiratory illnesses each year.⁴⁶ Novel mutations and reassortments of the virus will inevitably lead to the next IAV pandemic; therefore, the use and development of therapeutics that target conserved host pathways, rather than the virus itself, hold promise to curtail the impact of viral infection. Moreover, a heterogeneous response to IAV with the same virulence exists within the population, suggesting that host factors play a crucial role regulating the host response and determining the severity of lung injury.^{2,3,47} Additionally, experimental evidence from human studies and animal models of severe IAV show that viral titers do not always correlate with severity of disease, but rather ARDS induced “cytokine storm” is the major driver of morbidity and mortality.^{4,28,38,48}

Severe IAV infection is associated with inflammatory cytokines in humans and mice. Due to their pleiotropic and redundant effects, targeting of individual cytokines may not be a suitable approach to reduce pathology during IAV infection. Instead, dampening of the immune response may be more effective, as was the case with LUBAC destabilization noted above.³⁶ FDA-approved anti-inflammatory drugs, including corticosteroids and statins, have been proposed for the treatment of “cytokine storm” associated with severe IAV infection.⁴⁹ Moreover, current data regarding their efficacy is limited to mouse models and retrospective patient observations.^{49,50} Corticosteroids have been shown to be effective in limiting the inflammation in some lung pathologies.^{49–51} However, observational studies of the impact of corticosteroid treatment of IAV-infected patients suggest against their use; with administration associated with higher incidence of hospital-acquired pneumonia, longer duration of mechanical ventilation, and increased mortality.⁵⁰ Similarly, clinical evidence does not support corticosteroid treatment for COVID-19 lung

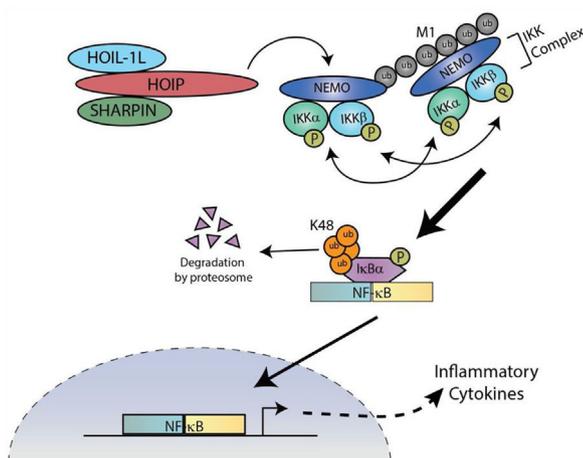


Fig. 3. LUBAC is necessary for NF- κ B dependent inflammation. LUBAC covalently attaches linear ubiquitin chains to NEMO, which facilitates the recruitment of additional IKK complexes. Stably docked IKK complexes result in the efficient transautophosphorylation and activation of proximal IKK α/β , followed by the phosphorylation and degradation of I κ B α . NF- κ B translocates to the nucleus to stimulate transcription of inflammatory genes.

injury.⁵² Statins are another class of drugs recognized for their ability to dampen inflammation.⁴⁹ While experimental evidence from mouse models using statins during IAV infection have been inconclusive regarding their benefit,^{49,53,54} retrospective analysis of patient data suggests an association between statin treatment and lower IAV mortality rates.⁵⁵ These examples highlight the need for new avenues of drug discovery and validations, as no currently available immune modulators have convincingly demonstrated their ability to improve outcomes during influenza infection.

LUBAC represents a potential new target for limiting the pathological inflammation that occurs during IAV infection. Several chemical inhibitors as well as peptides that bind HOIP have been used to inhibit LUBAC activity in cell culture^{56–59} and *in vitro* assays^{60,61} and support the specific targetability of LUBAC (Fig. 4). Currently, LUBAC inhibitors fall into two categories: those that target the catalytic activity of HOIP (*i.e.* BAY117082, Gliotoxin, HOIPIN) or those that disrupt the interaction between LUBAC components to destabilize the complex (*i.e.* stapled peptides). BAY117082, a small molecule commonly used as an inhibitor of NF- κ B activation. It has been observed that treatment of RAW 264.7 macrophages with BAY117082 prevented IL-1 stimulated formation of linear ubiquitin chains. Further investigation revealed that BAY117082 irreversibly inhibits LUBAC through a chemical reaction with cysteine residues in the active site of HOIP, the catalytic unit of LUBAC.⁵⁸ While BAY117082 represents a potent inhibitor of LUBAC activity, it targets multiple components of the ubiquitin system, including inhibition of E2 ubiquitin conjugating enzymes, and possible proteasome inhibition.⁵⁸ As such, use of BAY117082 is not suitable for the study of LUBAC-dependent physiological functions or therapeutic targeting of LUBAC activity in disease. Gliotoxin, a fungal metabolite, was identified in a high-throughput screening for LUBAC inhibitors using a time-resolved FRET-based screening system. While gliotoxin is known to have multiple cellular targets, it is able to inhibit LUBAC activity and downstream activation of NF- κ B at 10x lower concentrations.⁶¹ Gliotoxin's strong, irreversible binding to the catalytic site of HOIP makes it a selective inhibitor of LUBAC activity.⁶¹ Interestingly, the potency of gliotoxin has been shown to vary between cell types, with myeloid and lymphoid cells being more sensitive to gliotoxin-mediated NF- κ B inhibition than epithelial cells.^{61,62} However, this irreversible inhibition may quench the inflammatory response and increase susceptibility to secondary infections. HOIPINs are synthetic small molecules that reversibly inhibit LUBAC through targeting of HOIP activity, displaying both LUBAC specificity as well as low cytotoxicity. Sev-

eral derivatives have been made with varying degrees of efficacy (HOIPIN-1-8), with HOIPIN-8 showing significantly enhanced ability to prevent LUBAC-mediated NF- κ B activation in response to TNF- α without cytotoxicity compared to the other derivatives *in vitro*.⁵⁷ Conversely, stapled α -helical peptides developed based on specific LUBAC structures disrupt interactions necessary for stable complex formation.^{56,63} Stapled peptides are a class of synthetic macrocycles where the secondary α -helix structure is stabilized by the introduction of a hydrophobic bridge or "staple" that rigidifies specific areas to inhibit protein:protein interactions.⁵⁶ Stapled peptides based on the HOIP ubiquitin binding domain have been shown to successfully inhibit LUBAC activity *in vitro* by disrupting its interaction with HOIL-1L and destabilizing the overall complex.^{56,63} While several inhibitors of LUBAC have been developed and shown promise *in vitro*, no data is available detailing their efficacy *in vivo*. Further investigation in to these compounds which target LUBAC stability to modulate the degree of LUBAC activity is warranted as they may be therapeutically beneficial for the treatment of hyper-inflammatory response during viral infection, where a graded host response is necessary.

Conclusions

Within the past 150 years, IAV has been the causative agent of at least five pandemics (1889, 1918, 1957, 1968, 2009).^{47,64} In addition to IAV, novel viral threats, such as the coronavirus outbreaks of 2003, 2015 and 2019, quickly spread worldwide before virus-specific vaccines or pharmacological options could be developed. Thus, therapies that target conserved host pathways may provide a novel universal treatment strategies, regardless of viral sequence. Findings from animal models of IAV infection have shown that inhibition of the exuberant host immune response is associated with reduced lung injury and improved survival.^{11,14,27,28} However, current FDA-approved anti-inflammatory drugs, such as corticosteroids and statins, have failed to show benefit during severe IAV infection.^{49,50} Beyond viral infections, the amplitude of the inflammatory response has been shown to be a critical determinant in outcome during bacterial sepsis in community acquired pneumonia, a common complication post IAV infection.^{51,65} Analysis of a cohort of patients showed that reductions in the inflammatory response during bacterial pneumonia, both due to host inability to mount a response or the administration of anti-inflammatory steroids, lead to increased mortality.^{51,65} These clinical observations highlight the need to balance the inflammatory response during viral infection, not only to improve lung injury during the primary viral infection, but also to prevent poor outcomes to secondary infections.

In addition to the seasonal threat of influenza, we must also be cautious in the regulation of inflammation in treatment of the ongoing COVID-19 pandemic. While a subgroup of patients with severe COVID-19 develop 'cytokine storm',^{15,16,18} it must be remembered that current anti-inflammatory drugs have pleiotropic effects and lack the specificity needed to carefully calibrate the host response. As such, newly developed pharmacologics such as those that target LUBAC, a molecular rheostat of inflammatory signaling, have the potential to fine tune inflammation and moderate the host response. Further investigation of compounds which modulate LUBAC activity is warranted, as they may be therapeutically beneficial for the treatment of hyper-inflammatory response during viral infections, where a milder host response should improve outcomes.

Conflict of interest

The authors have declared that no conflict of interest exists.

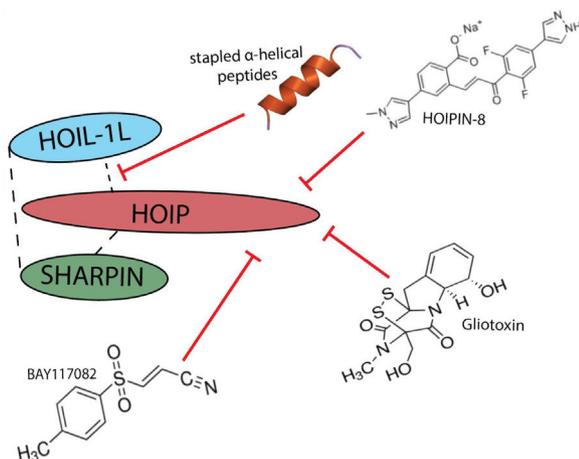


Fig. 4. Schematic representation of the small molecule inhibitors (BAY117082, Gliotoxin, HOIPIN-8) that target the catalytic domain of HOIP and the stapled α -helical peptides which disrupt the interaction between LUBAC components to destabilize the complex.

References

- Blank R, Napolitano LM. Epidemiology of ARDS and ALI. *Crit Care Clin*. 2011;27:439–58.
- Napolitano LM, Park PK, Raghavendran K, Bartlett RH. Nonventilatory strategies for patients with life-threatening 2009 H1N1 influenza and severe respiratory failure. *Crit Care Med*. 2010;38 4 Suppl:e74–90.
- Rothberg MB, Haessler SD. Complications of seasonal and pandemic influenza. *Crit Care Med*. 2010;38 4 Suppl:e91–7.
- Short KR, Kroeze E, Fouchier RAM, Kuiken T. Pathogenesis of influenza-induced acute respiratory distress syndrome. *Lancet Infect Dis*. 2014;14:57–69.
- Matute-Bello G, Downey G, Moore BB, Groshong SD, Matthay MA, Slutsky AS, et al. An official American Thoracic Society workshop report: features and measurements of experimental acute lung injury in animals. *Am J Respir Cell Mol Biol*. 2011;44:725–38.
- Cochi SE, Kempker JA, Annangi S, Kramer MR, Martin GS. Mortality Trends of Acute Respiratory Distress Syndrome in the United States from 1999 to 2013. *Ann Am Thorac Soc*. 2016;13:1742–51.
- Calfee CS, Delucchi K, Parsons PE, Thompson BT, Ware LB, Matthay MA, et al. Subphenotypes in acute respiratory distress syndrome: latent class analysis of data from two randomised controlled trials. *Lancet Respir Med*. 2014;2:611–20.
- Vadasz I, Sznajder JI. Gas Exchange Disturbances Regulate Alveolar Fluid Clearance during Acute Lung Injury. *Front Immunol*. 2017;8:757.
- Mauad T, Hajjar LA, Callegari GD, da Silva LF, Schout D, Galas FR, et al. Lung pathology in fatal novel human influenza A (H1N1) infection. *Am J Respir Crit Care Med*. 2010;181:72–9.
- Walsh KB, Teijaro JR, Wilker PR, Jatzek A, Fremgen DM, Das SC, et al. Suppression of cytokine storm with a sphingosine analog provides protection against pathogenic influenza virus. *Proc Natl Acad Sci U S A*. 2011;108:12018–23.
- Lin KL, Suzuki Y, Nakano N, Ramsburg E, Gunn MD. CCR2+ monocyte-derived dendritic cells and exudate macrophages produce influenza-induced pulmonary immune pathology and mortality. *J Immunol*. 2008;180:2562–72.
- Arankalle VA, Lole KS, Arya RP, Tripathy AS, Ramdasi AY, Chadha MS, et al. Role of host immune response and viral load in the differential outcome of pandemic H1N1 (2009) influenza virus infection in Indian patients. *PLoS One*. 2010;5:10.
- de Jong MD, Simmons CP, Thanh TT, Hien VM, Smith GJ, Chau TN, et al. Fatal outcome of human influenza A (H5N1) is associated with high viral load and hypercytokinemia. *Nat Med*. 2006;12:1203–7.
- Kobasa D, Jones SM, Shinya K, Kash JC, Copps J, Ebihara H, et al. Aberrant innate immune response in lethal infection of macaques with the 1918 influenza virus. *Nature*. 2007;445:319–23.
- Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan* China. *Lancet*. 2020;395(10223):497–506.
- Ruan Q, Yang K, Wang W, Jiang L, Song J. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan* China. *Intensive Care Med*. 2020.
- Richardson P, Griffin I, Tucker C, Smith D, Oechsle O, Phelan A, et al. Baricitinib as potential treatment for 2019-nCoV acute respiratory disease. *Lancet*. 2020;395(10223):e30–1.
- Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ, et al. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet*. 2020;395(10229):1033–4.
- Siren J, Imaizumi T, Sarkar D, Pietila T, Noah DL, Lin R, et al. Retinoic acid inducible gene-1 and mda-5 are involved in influenza A virus-induced expression of antiviral cytokines. *Microbes Infect*. 2006;8:2013–20.
- Ohman T, Rintahaka J, Kalkkinen N, Matikainen S, Nyman TA. Actin and RIG-I/MAVS signaling components translocate to mitochondria upon influenza A virus infection of human primary macrophages. *J Immunol*. 2009;182:5682–92.
- Kato H, Sato S, Yoneyama M, Yamamoto M, Uematsu S, Matsui K, et al. Cell type-specific involvement of RIG-I in antiviral response. *Immunity*. 2005;23:19–28.
- Davidson S, Crotta S, McCabe TM, Wack A. Pathogenic potential of interferon alpha in acute influenza infection. *Nat Commun*. 2014;5:3864.
- Whitsett JA, Alenghat T. Respiratory epithelial cells orchestrate pulmonary innate immunity. *Nat Immunol*. 2015;16:27–35.
- Pulendran B, Maddur MS. Innate immune sensing and response to influenza. *Curr Top Microbiol Immunol*. 2015;386:23–71.
- Hashimoto Y, Moki T, Takizawa T, Shiratsuchi A, Nakanishi Y. Evidence for phagocytosis of influenza virus-infected, apoptotic cells by neutrophils and macrophages in mice. *J Immunol*. 2007;178:2448–57.
- Hwang I, Scott JM, Kakarla T, Duriancik DM, Choi S, Cho C, et al. Activation mechanisms of natural killer cells during influenza virus infection. *PLoS One*. 2012;7:e51858.
- Oldstone MBA, Teijaro JR, Walsh KB, Rosen H. Dissecting influenza virus pathogenesis uncovers a novel chemical approach to combat the infection. *Virology*. 2013;435:92–101.
- Brazee PL, Soni PN, Tokhtaeva E, Magnani N, Yemelyanov A, Perlman HR, et al. FXYD5 Is an Essential Mediator of the Inflammatory Response during Lung Injury. *Front Immunol*. 2017;8:623.
- Herold S, Steinmueller M, von Wulffen W, Cakarova L, Pinto R, Pleschka S, et al. Lung epithelial apoptosis in influenza virus pneumonia: the role of macrophage-expressed TNF-related apoptosis-inducing ligand. *J Exp Med*. 2008;205:3065–77.
- Dawson TC, Beck MA, Kuziel WA, Henderson F, Maeda N. Contrasting effects of CCR5 and CCR2 deficiency in the pulmonary inflammatory response to influenza A virus. *Am J Pathol*. 2000;156:1951–9.
- Zhou G, Juang SW, Kane KP. NK cells exacerbate the pathology of influenza virus infection in mice. *Eur J Immunol*. 2013;43:929–38.
- Dove KK, Kleit RE. RING-between-RINGS—keeping the safety on loaded guns. *EMBO J*. 2012;31:3792–4.
- Ikeda F, Deribe YL, Skanland SS, Stieglitz B, Grabbe C, Franz-Wachtel M, et al. SHARPIN forms a linear ubiquitin ligase complex regulating NF-kappaB activity and apoptosis. *Nature*. 2011;471:637–41.
- Tokunaga F, Iwai K. LUBAC, a novel ubiquitin ligase for linear ubiquitination, is crucial for inflammation and immune responses. *Microbes Infect*. 2012;14(7–8):563–72.
- Brazee P, Dada LA, Sznajder JI. Role of Linear Ubiquitination in Health and Disease. *Am J Respir Cell Mol Biol*. 2016;54:761–8.
- Brazee PL, Morales-Nebreda L, Magnani ND, Garcia JG, Misharin AV, Ridge KM, et al. Linear ubiquitin assembly complex regulates lung epithelial-driven responses during influenza infection. *J Clin Invest*. 2020;130:1301–14.
- Vareille M, Kieninger E, Edwards MR, Regamey N. The airway epithelium: soldier in the fight against respiratory viruses. *Clin Microbiol Rev*. 2011;24:210–29.
- Guillot L, Nathan N, Tabary O, Thouvenin G, Le Rouzic P, Corvol H, et al. Alveolar epithelial cells: master regulators of lung homeostasis. *Int J Biochem Cell Biol*. 2013;45:2568–73.
- Sznajder JI. Alveolar Edema Must Be Cleared for the Acute Respiratory Distress Syndrome Patient to Survive. *Am J Respir Crit Care Med*. 2001;163:1293–4.
- Haas TL, Emmerich CH, Gerlach B, Schmukle AC, Cordier SM, Rieser E, et al. Recruitment of the linear ubiquitin chain assembly complex stabilizes the TNF-R1 signaling complex and is required for TNF-mediated gene induction. *Mol Cell*. 2009;36:831–44.
- Kenschke T, Tokunaga F, Ikeda F, Goto E, Iwai K, Dikic I. Analysis of nuclear factor-kappaB (NF-kappaB) essential modulator (NEMO) binding to linear and lysine-linked ubiquitin chains and its role in the activation of NF-kappaB. *J Biol Chem*. 2012;287:23626–34.
- Tokunaga F, Sakata S, Saeki Y, Satomi Y, Kirisako T, Kamei K, et al. Involvement of linear polyubiquitylation of NEMO in NF-kappaB activation. *Nat Cell Biol*. 2009;11:123–32.
- Smit JJ, Monteferrario D, Noordermeer SM, van Dijk WJ, van der Reijden BA, Sixma TK. The E3 ligase HOIP specifies linear ubiquitin chain assembly through its RING-IBR-RING domain and the unique LDD extension. *EMBO J*. 2012;31:3833–44.
- Dawood FS, Juliano AD, Reed C, Meltzer MI, Shay DK, Cheng PY, et al. Estimated global mortality associated with the first 12 months of 2009 pandemic influenza A H1N1 virus circulation: a modelling study. *Lancet Infect Dis*. 2012;12:687–95.
- CDC. 2009 H1N1 Pandemic Timeline. May 8, 2019 [cited 2020].
- Iuliano AD, Roguski KM, Chang HH, Muscatello DJ, Palekar R, Tempia S, et al. Estimates of global seasonal influenza-associated respiratory mortality: a modelling study. *Lancet*. 2018;391(10127):1285–300.
- Jain S, Kamimoto L, Bramley AM, Schmitz AM, Benoit SR, Louie J, et al. Hospitalized patients with 2009 H1N1 influenza in the United States. April–June 2009. *N Engl J Med*. 2009;361:1935–44.
- Short KR, Kasper J, van der Aa S, Andeweg AC, Zaaraoui-Boutahar F, Goeynber M, et al. Influenza virus damages the alveolar barrier by disrupting epithelial cell tight junctions. *Eur Respir J*. 2016;47:954–66.
- Darwish I, Mubareka S, Liles WC. Immunomodulatory therapy for severe influenza. *Expert Rev Anti Infect Ther*. 2011;9:807–22.
- Nedel WL, Nora DG, Salluh JJ, Lisboa T, Povoia P. Corticosteroids for severe influenza pneumonia: A critical appraisal. *World J Crit Care Med*. 2016;5:89–95.
- Antcliffe DB, Burnham KL, Al-Beidh F, Santhakumaran S, Brett SJ, Hinds CJ, et al. Transcriptomic Signatures in Sepsis and a Differential Response to Steroids. From the VANISH Randomized Trial. *Am J Respir Crit Care Med*. 2019;199:980–6.
- Russell CD, Millar JE, Baillie JK. Clinical evidence does not support corticosteroid treatment for 2019-nCoV lung injury. *Lancet*. 2020;395(10223):473–5.
- Kumaki Y, Morrey JD, Barnard DL. Effect of statin treatments on highly pathogenic avian influenza H5N1, seasonal and H1N1pdm09 virus infections in BALB/c mice. *Future Virol*. 2012;7:801–18.
- Radigan KA, Ulrich D, Misharin AV, Chiarella SE, Soberanes S, Gonzalez A, et al. The effect of rosuvastatin in a murine model of influenza A infection. *PLoS One*. 2012;7:35788.
- Vandermeer ML, Thomas AR, Kamimoto L, Reingold J, Gershman K, Meek J, et al. Association between use of statins and mortality among patients hospitalized with laboratory-confirmed influenza virus infections: a multistate study. *J Infect Dis*. 2012;205:13–9.
- Aguilar-Alonso F, Whiting AL, Kim YJ, Bernal F. Biophysical and biological evaluation of optimized stapled peptide inhibitors of the linear ubiquitin chain assembly complex (LUBAC). *Bioorg Med Chem*. 2018;26:1179–80.
- Katsuya K, Oikawa D, Iio K, Obika S, Hori Y, Urashima T, et al. Small-molecule inhibitors of linear ubiquitin chain assembly complex (LUBAC). HOIPINs, suppress NF-kappaB signaling. *Biochem Biophys Res Commun*. 2019;509:700–6.
- Strickson S, Campbell DG, Emmerich CH, Knebel A, Plater L, Ritorto MS, et al. The anti-inflammatory drug BAY 11-7082 suppresses the MyD88-dependent signalling network by targeting the ubiquitin system. *Biochem J*. 2013;451:427–37.
- Yang Y, Schmitz R, Mitala J, Whiting A, Xiao W, Ceribelli M, et al. Essential role of the linear ubiquitin chain assembly complex in lymphoma revealed by rare germline polymorphisms. *Cancer Discov*. 2014;4:480–93.
- Johansson H, Isabella Tsai YC, Fantom K, Chung CW, Kumper S, Martino L, et al. Fragment-Based Covalent Ligand Screening Enables Rapid Discovery of Inhibitors for the RBR E3 Ubiquitin Ligase HOIP. *J Am Chem Soc*. 2019;141:2703–10.

61. Sakamoto H, Egashira S, Saito N, Kirisako T, Miller S, Sasaki Y, et al. Gliotoxin suppresses NF- κ B activation by selectively inhibiting linear ubiquitin chain assembly complex (LUBAC). *ACS Chem Biol*. 2015;10:675–81.
62. Fitzpatrick LR, Wang J, Le T. In vitro and in vivo effects of gliotoxin, a fungal metabolite: efficacy against dextran sodium sulfate-induced colitis in rats. *Dis Sci*. 2000;45:2327–36.
63. Fujita H, Tokunaga A, Shimizu S, Whiting AL, Aguilar-Alonso F, Takagi K, et al. Cooperative Domain Formation by Homologous Motifs in HOIL-1L and SHARPIN Plays A Crucial Role in LUBAC Stabilization. *Cell Rep*. 2018;23:1192–204.
64. Taubenberger JK, Morens DM. The pathology of influenza virus infections. *Annu Rev Pathol*. 2008;3:499–522.
65. Davenport EE, Burnham KL, Radhakrishnan J, Humburg P, Hutton P, Mills TC, et al. Genomic landscape of the individual host response and outcomes in sepsis: a prospective cohort study. *Lancet Respir Med*. 2016;4:259–71.