

Review Article

ARCHIVOS DE Bronconeumología



www.archbronconeumol.org

Infant Bronchiolitis Endotypes and the Risk of Developing Childhood Asthma: Lessons From Cohort Studies



Heidi Makrinioti^{a,*}, Zhaozhong Zhu^a, Sejal Saglani^b, Carlos A. Camargo^a, Kohei Hasegawa^a

^a Department of Emergency Medicine, Massachusetts General Hospital, Harvard Medical School, Boston, MA, USA ^b National Heart and Lung Institute, Imperial College, London, United Kingdom

ARTICLE INFO

Article history: Received 28 December 2023 Accepted 14 February 2024 Available online 19 February 2024

Keywords: Asthma Bronchiolitis Endotype Epigenomics Genomics Metabolomics Microbiome Transcriptomics Proteomics

ABSTRACT

Severe bronchiolitis (i.e., bronchiolitis requiring hospitalization) during infancy is a heterogeneous condition associated with a high risk of developing childhood asthma. Yet, the exact mechanisms underlying the bronchiolitis-asthma link remain uncertain. Birth cohort studies have reported this association at the population level, including only small groups of patients with a history of bronchiolitis, and have attempted to identify the underlying biological mechanisms. Although this evidence has provided valuable insights, there are still unanswered questions regarding severe bronchiolitis-asthma pathogenesis. Recently, a few bronchiolitis cohort studies have attempted to answer these questions by applying unbiased analytical approaches to biological data. These cohort studies have identified novel bronchiolitis subtypes (i.e., endotypes) at high risk for asthma development, representing essential and enlightening evidence. For example, one distinct severe respiratory syncytial virus (RSV) bronchiolitis endotype is characterized by the presence of Moraxella catarrhalis and Streptococcus pneumoniae, higher levels of type I/II IFN expression, and changes in carbohydrate metabolism in nasal airway samples, and is associated with a high risk for childhood asthma development. Although these findings hold significance for the design of future studies that focus on childhood asthma prevention, they require validation. However, this scoping review puts the above findings into clinical context and emphasizes the significance of future research in this area aiming to offer new bronchiolitis treatments and contribute to asthma prevention.

© 2024 SEPAR. Published by Elsevier España, S.L.U. All rights reserved.

Introduction

Infant bronchiolitis is the most common lower respiratory infection and the leading cause of infant hospitalizations in the U.S.¹ Severe bronchiolitis (i.e., bronchiolitis requiring hospitalization) is also a major risk factor for the development of childhood asthma.^{2,3} This bronchiolitis-asthma association is described by both birth cohort studies and bronchiolitis cohort studies.^{4,5} Birth cohort studies report that, at the population level, infants with a history of severe bronchiolitis are at higher risk of childhood asthma.^{4,6} Likewise, cohort studies of severe bronchiolitis demonstrate that approximately 30% of infants with severe bronchiolitis develop asthma by age 6–7 years.^{2,7} Yet, the exact mechanisms underlying the bronchiolitis-asthma link remain uncertain.

While bronchiolitis has long been considered a single disease entity,⁸ recent cohort studies describe clinical phenotypes of bronchiolitis that are associated with increased risk for childhood

* Corresponding author. E-mail address: cmakrynioti@mgh.harvard.edu (H. Makrinioti). asthma.^{2,5,9–11} For example, infants hospitalized with rhinovirus bronchiolitis with a history of previous wheezing episodes and eczema have a higher risk of developing asthma compared to those with RSV infection.^{2,5,12} However, the bronchiolitis-asthma link involves a complex interplay among multi-level factors, such as respiratory viruses and non-virus exposures (i.e., air pollution, climate exposures), host genetics, and immune responses.^{13–15} Therefore, it is essential to gain a deeper understanding of these intricate interrelationships.

Toward this direction, recent studies have utilized unbiased and integrated (i.e., clustering) approaches in the analysis of multi-level omics data (e.g., the genome, epigenome, transcriptome, proteome, metabolome, microbiome, metatranscriptome) from nasal airway samples in addition to peripheral blood samples.^{16–19} These emerging findings help us understand that asthma susceptibility is not defined solely by genetic variation at a single locus (e.g., *17q21* locus)—a finding that derives from a single omics approach.^{20–22} A more plausible framework involves, for example, genetically driven metabolites and the genetic loci regulating these metabolites (e.g., sphingolipids with genetic variation at *1q32* locus) associated with the asthma risk, a finding that derives from an integrative genetic-

metabolomic approach.¹⁸ Importantly, clustering approaches have identified distinct bronchiolitis profiles (i.e., endotypes) at differential risks for childhood asthma.^{23,24} Bronchiolitis endotyping is important, as it is the most suitable research approach to address disease heterogeneity.

Despite the increasing research significance, bronchiolitis endotyping approaches leave us with the challenge of clinical translation (i.e., how to use the information for the development of targeted bronchiolitis treatments or of strategies for the primary prevention of asthma). Rising to this challenge would represent the beginning of precision medicine for a large population of children. This review focuses on the description of severe bronchiolitis endotypes at differential risks for development of childhood asthma. In addition, this review summarizes evidence deriving from nonclustering approaches in birth and bronchiolitis cohort studies with the aim to define susceptibility to childhood asthma.

Literature search

This is a scoping review discussing past and current evidence related to molecular (i.e., omics) profiles in the general population and in infants with severe bronchiolitis, examining associations with a high risk of asthma development. Relevant studies have been identified through query of the Medline, Embase, and Cochrane databases for English language articles published up to September 2023 using the terms "birth cohort", "bronchiolitis", "bronchiolitis cohort", "genomics", "epigenomics", "transcriptomics, "proteomics", "metabolomics", "microbiome", "omics", "endotype", and "asthma". Studies have been selected for discussion in this review based on topic relevance. Publications cited by articles identified through the search strategy have been included as appropriate.

Evidence review

Severe bronchiolitis in infancy and childhood asthma risk: what we know and which the knowledge gap is

For decades, bronchiolitis has been considered and managed as a single disease entity.^{8,25,26} However, it is recognized that severe bronchiolitis is heterogeneous in terms of both clinical presentation and the risk of chronic respiratory sequelae, such as recurrent wheeze and childhood asthma.^{2,27,28} Studies have identified several risk factors for subsequent asthma development by utilizing a single-level approach.^{29–31} For example, specific respiratory viruses (e.g., respiratory syncytial virus [RSV] or rhinovirus) have been consistently identified as risk factors for childhood asthma development.^{29,32–34} Despite the limited data, it is still possible to draw some insights- For example, infants with rhinovirus bronchiolitis have a 3 times higher risk of asthma compared to those with RSV bronchiolitis.³⁵ In addition, traffic-related air pollution or the development of early-life allergic sensitization are associated with an increased risk for childhood asthma.^{29,36}

While the identification of these single-level risk factors has advanced our understanding, they do not address the complex interplay between exposures at multiple levels (e.g., environmental exposures, host genome, transcriptome, metabolome, and microbiome) and how they may contribute to the bronchiolitis-asthma link.^{16,23}

How omics approaches can help us address the knowledge gap

Omics approaches enable the comprehensive characterization and analysis of biological molecules.³⁷ Therefore, these approaches provide valuable insights into the structure and function of human tissues, which aid further in the identification of molecular mechanisms underlying disease development that serve as potential therapeutic targets.^{28,37,38} Birth and bronchiolitis cohort studies from the U.S. (e.g., the Multicenter Airway Research Collaboration [MARC], RSV bronchiolitis in Early Life [RBEL], Infant Susceptibility to Pulmonary Infections and Asthma Following RSV Exposure [INSPIRE], Childhood Origins of Asthma [COAST]) and Europe (e.g., Breathing Together, Copenhagen Prospective Studies on Asthma [COPSAC] and Manchester Asthma and Allergy study [MAAS]) utilize mainly non-clustering and, few of them, clustering omics approaches (i.e., endotyping) with the aim to examine the mechanisms that underlie the development of childhood asthma.^{17,39–44} To date, most severe bronchiolitis endotyping data derive from the MARC-35 cohort study.^{13,17,23,24,45} MARC-35 is severe bronchiolitis cohort that completed enrolment of 1016 hospitalized infants across 17 sites in the US. In this diverse cohort (53% African American or Hispanic), investigators collected biospecimens, including nasal swabs, nasopharyngeal aspirates, and peripheral blood samples at the index hospitalization.⁹ The following sections discuss available evidence from both non-clustering and clustering omics approaches in birth and bronchiolitis cohort studies.

Omics approaches in birth cohort studies

U.S. and European birth cohort studies have advanced our understanding around childhood asthma etiology by using nonclustering (i.e., non-grouping) omics (e.g., genomic, epigenomic, metabolomic, microbiome) approaches. These studies do not focus solely on infants with severe bronchiolitis. Representative findings from these studies, including types of samples used and methodology (e.g., omics approach) are discussed below and are summarized in Table 1.

Genetics approaches

Of genetic variants associated with the asthma risk, genomewide association studies (GWAS) and candidate-gene association studies link those in orosomucoid-like sphingolipid biosynthesis regulator 3 (ORMDL3) in chromosome 17q21 to development of childhood asthma.46,47 Other commonly reported loci in the association with asthma include, for example, the Denn domain containing 1B (DENND1B), interleukin (IL) 1 receptor ligand 1(IL1RL1), RAD50, cadherin-related family member 3 (CDHR3), and IL33 loci.⁴⁸ These loci are also associated with a higher risk for a childhood asthma phenotype characterized by a high number of virus-induced exacerbations.⁴⁹ Additionally, a Danish GWAS reported an interaction between CDHR3 and gasdermin B (GSDMB) loci in the development of early childhood asthma.⁵⁰ Taken together, these findings, albeit not deriving from infants with severe bronchiolitis, demonstrate the importance of understanding genetic heterogeneity in childhood asthma and its phenotypes.⁵¹

Epigenomic approaches

We next summarize epigenomics evidence from non-clustering approaches. A consortium-based meta-analysis of epigenomewide association studies (EWAS) utilizing whole blood samples and identified hypomethylated 5'-C-phosphate-G-3' (CpG) sites that are associated with high asthma risk.⁵² These CpG sites and the associated transcriptional profiles indicate, for example, activation of eosinophils and reduction in naïve T cells as possible important mediators in the pathway linking environmental exposures at early years to childhood asthma development.⁵² Data from other EWAS using whole blood samples also report an epigenetically driven eosinophil activation pattern in the development of childhood asthma.^{53,54}

In addition to the identification of individual CpGs, a recent EWAS focuses on differentially methylated regions (DMRs) to

Table 1

Representative birth cohort studies describing associations of omics non-clustering approaches and endotypes with the development of childhood asthma.

Citation	Study type	Study population and size	Biological specimen source	Clustering vs non-clustering approaches and type of omics profiling	Pathophysiological mechanisms	Outcome(s)	Measurement of association
Çalışkan M. et al., 2013 ²⁰	Birth cohort study	497 infants recruited at the COAST and COPSAC birth cohort studies	Peripheral blood samples (SNPs in 17q21 gene)	Non-clustering approach Genomic profiling (microarray)	Interaction between 17q21 SNP genotypes and respiratory viruses	Incident asthma by seven years	17q21 genotypes in infants presenting later with rhinovirus-induced wheezing illness associated with increased risk for incident asthma by seven years OR: 5.2; 95% CI: 2.8–9.9; p < 0.001
Rosas-Salazar C. et al., 2016 ⁴⁰	Birth cohort study	<u>99 infants</u> with RSV-bronchiolitis 33 healthy controls	Nasopharyngeal airway samples (respiratory virus and nasopharyngeal microbiome)	Non-clustering approach Microbiome analysis (bacterial 16S rRNA gene sequencing)	Interrelationship between respiratory viruses and major bacteria species	Incident asthma by five years	Infants with RSV-bronchiolitis and high nasopharyngeal levels of <i>Streptococcus pneumoniae</i> , <i>Moraxella catarrhalis</i> , and <i>Haemophilus influenza</i> have an increased risk of developing asthma by five years p < 0.001
Hallmark B. et al., 2021 ⁷⁵	Birth cohort study	<u>3786 infants</u> from seven US birth cohort studies CREW network	Peripheral blood samples (SNPs in 17q21 gene)	Non-clustering approach Genomic profiling (microarray)	Interaction between 17q21 SNP genotypes, IgE levels, and clinical characteristics	Incident asthma by eleven years	17q21 genotypes in infants presenting with persistent wheezing illness compared to transient wheezing illness associated with increased risk for incident asthma by eleven years p < 0.001
Eliasen A. et al., 2022 ⁵⁰	Birth cohort study	6532 infants from COPSAC 2000 and COPSAC 2010 studies	Peripheral blood samples (SNPs in <i>CDHR3</i> and <i>GSDMB</i> genes)	Non-clustering approach Genomic profiling (microarray)	Interaction between SNPs in CDHR3 and GSDMB genes and peripheral blood IL17A levels during respiratory viral infections	Incident asthma by six years	Interaction between SNPs in CDHR3 and CSDMB genes and increased <i>IL17A</i> peripheral blood levels associated with increased risk for incident asthma risk by six years p < 0.001
Gürdeniz G. et al., 2022 ⁵⁹	Birth cohort study	<u>1064 infants</u> from COPSAC 2000 and COPSAC 2010 studies	Peripheral blood samples (dry blood spot) furan fatty acid profiles	Non-clustering approach Targeted metabolomics profiling (LC-MS)	Interaction between furan fatty acid profiles and clinical characteristics	Incident asthma by six years	The 3-carboxy-4-methyl-5- propyl-2-furan propanoic acid profile is associated with a decreased risk for incident asthma by six years HR = 0.89; <i>p</i> = 0.002

reduce the dimension of epigenome-wide methylation data.^{54,55} In this context, an epigenome-wide meta-analysis of associations between whole blood methylation sites and risk of school-age asthma identified 36 DMRs correlated with expression of genes that are already targeted in asthma treatments (e.g., *IL5R* gene, potassium voltage-gated channel subfamily H member [*KCNH*] gene).⁵⁴ The substantial impact of this research would be the use of CpGs and DMRs identified in birth cohort studies as potential therapeutic targets or biomarkers for subsequent asthma risk.^{56,57}

Metabolomic approaches

In addition to epigenomic profiles, an increasing number of scientific reports describe that the profiling of end products of cellular metabolism (i.e., metabolomics) is linked to the development of persistent wheezing and asthma in childhood.^{58–60} For example, recent findings from the COPSAC cohort using dried blood spots at birth demonstrate peripheral blood metabolomic profiles at higher risk for asthma development.⁵⁹ Specifically, decreased newborn tryptophan, bile acid, and phenylalanine metabolism were associated with a higher asthma risk.⁵⁹ In support of this evidence, bile metabolism pathways detected in plasma samples were also implicated in asthma pathogenesis following RSV bronchiolitis in U.S. birth cohort studies.⁶¹ Furthermore, the COPSAC₂₀₁₀ study has investigated whether a dietary intervention during pregnancy can decrease the risk of asthma.⁶⁰ Indeed, a randomized controlled trial of fish oil-derived n-3 long-chain polyunsaturated fatty acid supplements to pregnant mothers found that the intervention reduces the risk of incident asthma by age 5 years.⁶⁰ The study also found that infants born to mothers who received fish oil supplements and had reduced tryptophan-related metabolites and increased tyrosine- and glutamic acid-related metabolites were less likely to develop asthma by age 5 years.⁶⁰ In summary, birth cohort studies have identified that metabolome regulation in early life can be associated with risk of childhood asthma.

Microbiome approaches

In addition to metabolomic profiles, changes in the airway microbiome are considered to play a key role in the development of childhood asthma.^{62,63} These data derive from analysis of upper airway specimens since lower airway sampling (e.g., bronchoscopy) is invasive in young infants and usually takes place under absolute indications (e.g., intubation). In addition, studies have suggested that upper airway sampling possibly represents the lung transcriptome and microbiome profiles in children.^{64,65}

Firstly, findings from the COPSAC₂₀₁₀ cohort reported that reduced microbiome alpha diversity and an increase in relative abundance of *Veillonella* and *Prevotella* in the upper airway at age 1 month was associated with higher asthma development risk by age 6 years.⁶⁶ A higher relative abundance of these genera is also linked to upper airway immune profiles characterized by reduced levels of proinflammatory mediators (i.e., tumor necrosis factor- α [TNF- α] and IL-1 β).⁶⁶ In addition to these microbiome profiles, a U.S. birth cohort study also reported that detection of *Streptococcus pneumoniae* and *Moraxella catarrhalis* at the nasopharynx of infants with reported RSV-positive bronchiolitis was associated with increased asthma risk by age 6 years.⁴⁰

Furthermore, COPSAC cohort studies have identified that several factors are critical in determining the diversity of the upper airway and gut microbiome of children, and their association with asthma risk. These factors include the mode of delivery, the presence of older siblings, and the age gap to the closest older sibling.^{67,68} The above findings showcase that changes in airway microbiome in early life may be linked to the development of childhood asthma.

On the whole, birth cohort studies have enhanced our understanding around early-life origins of childhood asthma, despite not focusing on infants at high risk to develop childhood asthma (i.e., infants with severe bronchiolitis). The next section focuses on evidence from severe bronchiolitis cohort studies.

Omics approaches in severe bronchiolitis cohort studies

Reported omics data from both MARC-35 and other severe bronchiolitis cohorts⁶⁹ derive from both non-clustering (i.e., associations between severe bronchiolitis omics profiles and asthma development)^{14,18,70,71} and clustering (endotyping)^{13,17,23,24,45,72} approaches. Therefore, in this section and in Table 2, we summarize representative findings from both approaches.

Non-clustering approaches

Important findings regarding risk factors for the development of childhood asthma derive from the analysis of nasopharyngeal airway biological samples from infants with severe bronchiolitis. Specifically, an analysis of data from 244 infants with severe bronchiolitis in the MARC-35 cohort study confirmed the complexity of the interactions between the nasopharyngeal airway microbiome, host transcriptome, and metabolome in regard to the development of childhood asthma.¹⁴ Briefly, the investigators reported that an increased abundance of S. pneumoniae, particularly in infants with a non-rhinovirus infection, in combination with a downregulation of type I and II interferon (IFN) transcription, and an upregulation of T cell activation, were associated with an increased risk for asthma development by age six years. Another analysis described relevant findings in regard to IFN regulation, and further highlighted that RSV-rhinovirus coinfection, in correlation to an increased abundance of Haemophilus influenzae, and downregulation of type I IFN transcription, were associated with a higher risk for asthma development.⁷¹ In regard to whether the specific bacteria (e.g., S. pneumoniae) is a pivotal factor in these associations, there are now confirmatory findings in another US cohort.⁴⁰ However, the evidence around the causal role of these bacterial species (i.e., S. pneumoniae) in subsequent asthma development remains inconclusive. For example, an important unanswered question is whether these bacteria species merely acquire a pathogenic role in hosts who have already increased susceptibility to developing asthma.73

In addition to the respiratory virus, microbiome, and transcriptome findings, MARC-35 has also identified metabolomic profiles associated with a high risk for developing childhood asthma. An analysis performed for the entire cohort (i.e., 1016 infants with severe bronchiolitis), for example, links the upregulation in nasopharyngeal glutathione metabolism to a high risk of childhood asthma.⁷⁰

Another interesting approach in genomic and metabolomic data from the MARC-35 cohort indicates that researchers can utilize metabolomic profiles to further define risk for asthma development in individuals with known genetic susceptibility to asthma.¹⁸ More specifically, Ooka et al. identified significant single nucleotide polymorphisms (SNPs) within loci aligned but not definitely colocalized (i.e., colocalization posterior probability \geq 0.5) with known asthma-susceptibility genes (e.g., *ADORA1*, *MUC16*) being associated with dysregulation of metabolic pathways and high asthma risk.¹² These findings further underscore severe bronchiolitis heterogeneity investigated holistically through clustering approaches.

Clustering approaches (i.e., bronchiolitis endotyping)

The MARC-35 cohort is the first study identifying severe bronchiolitis endotypes at high risk for asthma development. These endotypes are derived from the analyses of nasopharyngeal or peripheral blood samples or both sample types. First, there are several MARC-35 studies defining severe bronchiolitis endotypes by utilizing nasopharyngeal airway samples.^{17,19,23,24,45,72} The most

Table 2

Representative bronchiolitis cohort studies describing associations between omics endotypes and the development of childhood asthma.

Citation	Study type	Study population and size	Biological specimen source	Clustering vs non-clustering approaches and type of omics profiling	Pathophysiological mechanisms	Outcome(s)	Measurement of association
Raita Y. et al., 2021 ²³	Bronchiolitis cohort study	<u>122 infants</u> with rhinovirus-induced severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasopharyngeal airway samples (rhinovirus subtypes, bacteria species of the nasopharyngeal microbiome, metabolome, and type 2 cytokines levels)	Clustering approach Metabolomic, respiratory viruses, and proteins profile	Interrelationship between rhinovirus species, major bacteria species, metabolomic profiles, and type 2 cytokines in the nasopharyngeal airway	Incident asthma by five years	Infants with endotype (atopic, rhinovirus C-positive, high nasopharyngeal levels of <i>Moraxella catarrhalis</i> and increased levels of type 2 cytokines) have a higher risk to develop asthma by five years OR: 3.74; 95% CI: 1.21–12.6; p = 0.03
Raita Y. et al., 2021 ¹⁷	Bronchiolitis cohort study	221 infants with RSV-induced severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasopharyngeal airway samples (coinfection with rhinovirus, microbiome, metabolome, and transcriptome)	Clustering approach Metabolomic, transcriptomic, respiratory virus, and bacteria profile	Interrelationship between RSV infection, RSV and rhinovirus double infection, major bacteria species, metabolomic profiles, and immune mediators' levels	Incident asthma by five years	Infants with endotype (atopic, nasopharyngeal high levels of <i>Streptococcus pneumoniae</i> , <i>Moraxella catarrhalis</i> , and increased <i>IFN-α</i> and - γ gene transcription) have a higher risk to develop asthma by five years OR: 6.00; 95% CI: 2.08–21.9; p = 0.002
Fujiogi M. et al., 2022 ⁴⁵	Bronchiolitis cohort study	<u>917 infants</u> with severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasopharyngeal airway samples (respiratory virus type, nasopharyngeal airway transcriptome, metabolome, and lipidome)	Clustering approach Transcriptomic, metabolomic, and lipidomic profiles	Interrelationship between respiratory viruses, nasopharyngeal airway transcriptomic, metabolomic, and lipidomic profiles	Incident asthma by six years	Infants with endotype (atopic, higher proportion of rhinovirus infection, IgE sensitization, low sphingolipids, and lower abundance of polyunsaturated fatty acids) have a higher risk to develop asthma by six years unadj OR: 3.60; 95% CI: 2.31–5.62; p < 0.001
Raita Y. et al., 2022 ⁷¹	Bronchiolitis cohort study	244 infants with severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasopharyngeal airway samples (respiratory virus type, nasopharyngeal airway transcriptome, and metatranscriptome)	Non-clustering approach Transcriptomic and metatranscriptomic profiles	Interrelationship between respiratory viruses, nasopharyngeal airway transcriptomic, and metatranscriptomic profiles	Incident asthma by six years	Infants with endotype B (RSV-rhinovirus positive, higher levels of <i>Haemophilus</i> <i>influenzae</i> , downregulated type I IFN pathways, upregulated T helper-17 pathways) have a higher risk of developing asthma by six years adj OR: 2.81; 95% CI: 1.11-7.26; p < 0.05
Zhu Z. et al., 2022 ¹⁴	Bronchiolitis cohort study	244 infants with severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasopharyngeal airway samples (respiratory virus type, nasopharyngeal airway transcriptome, metabolome, and metatranscriptome)	Non-clustering approach Transcriptomic, metatranscriptomic, and metabolomic profiles	Interrelationship between respiratory viruses, nasopharyngeal airway transcription, metatranscriptomic, and metabolomic profiles	Incident asthma by six years	Infants with rhinovirus-negative bronchiolitis, high levels of <i>Streptococcus pneumoniae</i> , lower abundance of fatty acid-related metabolites, and decreased type I and II IFN transcription are associated with a higher risk of developing asthma by six years FDR < 0.05 for transcriptome pathways and associations between <i>Streptococcus</i> <i>nneumoniae</i> and differentially

pneumoniae and differentially expressed transcripts

Table 2 (Continued)

Citation	Study type	Study population and size	Biological specimen source	Clustering vs non-clustering approaches and type of omics profiling	Pathophysiological mechanisms	Outcome(s)	Measurement of association
Zhu Z. et al., 2023 ⁷⁶	Bronchiolitis cohort study	575 infants with severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasal miRNA (respiratory virus type, differentially expressed miRNAs detected in nasal swab samples)	Non-clustering approach	Interrelationship between respiratory viruses, nasal airway miRNA, and mRNA levels	Incident asthma by six years	Differentially expressed miRNAs were negatively correlated with their mRNA targets (e.g., hsa-miR-324-3p/IL13), which were enriched in asthma-related pathways FDR < 0.05
Zhu Z. et al., 2022 ²⁴	Bronchiolitis cohort study	<u>914 infants</u> with severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasopharyngeal airway samples (respiratory virus type, nasopharyngeal airway metabolome)	Clustering approach Transcriptomic, metabolomic, and respiratory virus profile	Interrelationship between respiratory virus type and nasopharyngeal metabolomic profiles	Incident asthma by six years	Infants with endotype (lower abundance of fatty acid-related metabolites, downregulated type I and II IFN) have a higher risk of developing asthma by six years adj OR: 2.22; 95% CI: 1.07–4.69; p < 0.05
Zhu Z. et al., 2023 ⁷⁶	Bronchiolitis cohort study	575 infants with severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasal miRNA (respiratory virus type, differentially expressed miRNAs detected in nasal swab samples)	Non-clustering approach	Interrelationship between respiratory viruses, nasal airway miRNA, and mRNA levels	Incident asthma by six years	Differentially expressed miRNAs were negatively correlated with their mRNA targets (e.g., hsa-miR-324-3p/IL13), which were enriched in asthma-related pathways FDR < 0.05
Fujiogi M. et al., 2021 ¹⁶	Bronchiolitis cohort study	<u>140 infants</u> with severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Nasopharyngeal airway and peripheral blood samples (infection with RSV or rhinovirus, nasopharyngeal and serum metabolome)	Clustering approach Nasopharyngeal, serum metabolomic, and respiratory virus profiles	Interrelationship between RSV and rhinovirus infection, nasopharyngeal and serum metabolomic profiles	Incident asthma by five years	Integrated upregulated nasopharyngeal-serum carnitine metabolic pathways associated with incident asthma by five years adj OR: 1.48; 95% CI: 1.11–1.99; p < 0.05
Kyo M. et al., 2022 ⁷⁰	Bronchiolitis cohort study	<u>1013 infants</u> with severe bronchiolitis <u>1013 infants</u> with nasopharyngeal metabolomic profiling data <u>140 infants</u> with serum metabolomic profiling data	Nasopharyngeal airway and peripheral blood samples (nasopharyngeal and serum glutathione-related metabolites)	Non-clustering approach Transcriptomic, metatranscriptomic, and respiratory virus profile	Correlations between respiratory viruses, nasopharyngeal metatranscriptome, and metabolomic profiles	Incident asthma by six years	Nasopharyngeal glutathione-related metabolic signature negatively correlated with incident asthma by six years adj OR: 0.90; 95%CI: 0.82–0.99; p=0.04

Table 2

(Continued)

Citation	Study type	Study population and size	Biological specimen source	Clustering vs non-clustering approaches and type of omics profiling	Pathophysiological mechanisms	Outcome(s)	Measurement of association
Torgerson D.G. et al., 2015 ⁷⁷	Bronchiolitis cohort study	206 infants with severe bronchiolitis from the RBEL study	Peripheral blood samples (gene variants)	Non-clustering approach Genomic profiling	Correlations between respiratory viruses and gene variants	Incident asthma by seven years	Single variants in <i>ADRB2</i> , <i>FLG</i> , and <i>NCAM1</i> genes in European Americans and <i>NOS1</i> in African Americans were associated with an increased risk for incident asthma by seven years $p = 4.6 \times 10^{-4}$, 1.9×10^{-13} , 5.0×10^{-5} , and 2.3×10^{-11} respectively
Törmänen S. et al., 2017 ⁷⁸	Bronchiolitis cohort study	<u>166 infants</u> with severe bronchiolitis	Peripheral blood samples (SNPs in Toll-like receptor genes)	Non-clustering approach Genomic profiling	Correlations between respiratory viruses and gene variants	Incident asthma by seven years	The variant <i>TLR10 rs4129009</i> and <i>TLR7 rs179008</i> are associated with an increased risk for incident asthma by seven years p < 0.05 for both
Ooka T. et al., 2022 ¹³	Bronchiolitis cohort study	<u>140 infants</u> with severe bronchiolitis randomly selected out of 921 infants from the MARC-35 cohort study	Peripheral blood and nasopharyngeal airway samples (serum proteome and respiratory virus data)	Clustering approach Proteomic and respiratory viruses profile	Interrelationship between respiratory viruses and serum proteome profiles	Incident asthma by six years	The endotype of atopic infants with rhinovirus-positive bronchiolitis and $NF\kappa B$ -related pathways at higher risk to develop asthma by six years adj OR: 4.04; 95% CI: 1.49–11.0; p =0.006
Shibata R. et al., 2023 ¹⁹	Bronchiolitis cohort study	<u>1016 infants</u> with severe bronchiolitis	Peripheral blood and nasopharyngeal airway samples (total lgE and respiratory virus data)	Clustering approach Respiratory viruses and total lgE levels	Interrelationship between respiratory viruses and total lgE levels	Incident asthma by six years	The cluster of infants with high total IgE, high RSV virus load, and low RV load) had a significantly higher risk of developing asthma by six years adj OR: 2.93; 95% CI: 1.02–8.43; <i>p</i> = 0.046

Abbreviations: ADRB2, beta-2 adrenergic receptor; CDHR3, cadherin-related family member 3; COAST, Childhood Origins of Asthma; COPSAC, Copenhagen Prospective Studies in Asthma in Childhood; CREW, Children's Respiratory and Environmental Workgroup; FLG, filaggrin; GSDMB, gasdermin B; HR, hazards ratio; IFN, interferon; IL17A, interleukin 17A; IgE, immunoglobulin E; LC–MS, liquid chromatography mass spectrometry; MARC, Multicenter Airway Research Collaboration; NCAM1, neural cell adhesion molecule 1; OR, odds ratio; SNP, single nucleotide polymorphism; RBEL, RSV bronchiolitis in early life study; TLR, toll-like receptor.

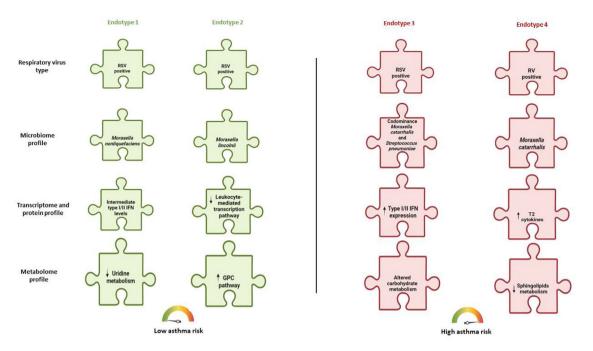


Fig. 1. Severe bronchiolitis endotypes at differential risks for developing childhood asthma This figure represents a collective pattern of four severe bronchiolitis endotypes, as defined through the analysis of nasopharyngeal airway samples, at differential risks for developing childhood asthma. Each puzzle piece represents a biological profile. There are four distinct rows of puzzle pieces linking to biological profiles: respiratory viruses, microbiome, transcriptome and protein, and metabolome profile. There are four distinct columns (i.e., four sets) of puzzle pieces that represent the four distinct endotypes derived from the compilation of these four biological profiles. Among the four sets of puzzle pieces, the two sets on the left side (in green color) represent endotypes at low risk for developing asthma, whilst the two other sets on the right side (in red color) represent endotype 1 represents RSV bronchiolitis with a high abundance of *Moraxella nonliquefaciens*, intermediate type I/II IFN levels, and downregulated GPC metabolism. Endotype 3 represents RSV bronchiolitis with a of *M. catarrhalis* increased levels of T2 cytokines (e.g., IL-4, IL-5, and IL-13), and downregulated sphingolipids metabolism. *Abbreviations*: GPC, glycerophosphocholine; IFN, interferor; RSV, respiratory syncytial virus; RV, rhinovirus; T2, type 2.

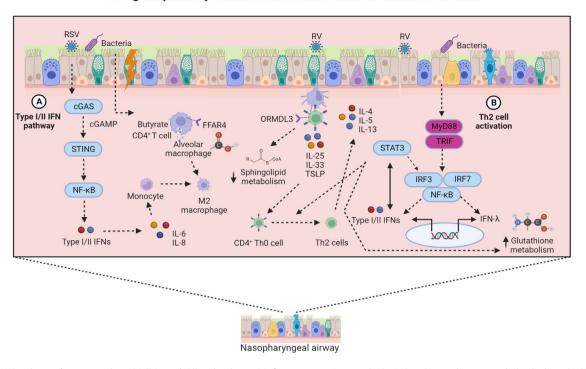
recent of these studies applied consensus clustering approaches to clinical, virus and nasopharyngeal airway lipidomic data and identified four lipidomic endotypes at differential risks for childhood asthma.⁴⁵ Out of these four endotypes, the endotype of infants with a higher proportion of parental asthma, IgE sensitization, rhinovirus infection and decreased sphingolipids metabolism was at greater asthma risk compared to other three endotypes.⁴⁵ Another MARC-35 study utilizing nasopharyngeal airway samples applied a consensus clustering approach to metabolomic and transcriptomic data.²⁴ This study identified five metabotypes at differential risks for childhood asthma. Out of these endotypes, the endotype of infants with increased nasopharyngeal amino acid metabolism and decreased fatty acid metabolism presented the highest risk for childhood asthma development. Clustering approaches were also applied in other omics data from the MARC-35 cohort.^{17,23} For example, the application of similarity network fusion approach to the nasopharyngeal airway microbiome, transcriptome and metabolome data, Raita et al. identified four nasopharyngeal endotypes in 221 infants with RSV bronchiolitis and another four endotypes in 122 infants with rhinovirus bronchiolitis.^{17,23} In RSV bronchiolitis, a high proportion of IgE sensitization and codominance of M. catarrhalis and S. Pneumoniae, in addition to increased type I/II IFN expression, were associated with the highest asthma risk.¹⁷ In rhinovirus bronchiolitis, an increased abundance of M. catarrhalis, upregulation of STAT3 transcription and type 2 cytokines were associated with the highest risk for asthma development.²³ These two representative bronchiolitis endotypes at high risk for childhood asthma are depicted in Figs. 1 and 2. Fig. 2 describes in detail possible pathobiological mechanisms that underlie these severe bronchiolitis endotypes.

In addition to these MARC-35 studies that utilize nasopharyngeal airway samples, another MARC-35 study examined peripheral serum samples for endotyping.¹³ Ooka et al. applied clustering approaches to integrate clinical, respiratory virus and serum proteome data in 140 infants with severe bronchiolitis from MARC-35. The study identified two endotypes at highest asthma risk; (A) the endotype of rhinovirus-positive, atopic infants with dysregulated nuclear factor kappa beta signaling pathways; and (B) the endotype of RSV- and rhinovirus-double positive non-atopic infants with dysregulated TNF- α signaling pathways.¹³

Closing section

Although the endotypes described above are associated with various degrees of risk for developing childhood asthma, some biological pathways have been shown to play a central role in the definition of more than one of these severe bronchiolitis endo-types (e.g., variability in *ORMDL3* gene and associated sphingolipid metabolism, type I and II IFN regulation, fatty acid and amino acid metabolism).^{17,23,46,59} Lastly, we recognize that the bronchiolitis endotypes in particular may not be mutually exclusive.

It is important to remember that MARC-35 is a severe bronchiolitis cohort. Therefore, attempts to draw direct comparisons between the MARC-35 cohort and other cohorts, such as the general birth cohorts, are challenging. However, some biological pathways have been shown to play a central role in the definition of more than one of these severe bronchiolitis endotypes. These pathways include the variability in *ORMDL3* gene and associated sphingolipid metabolism, type I and II IFN regulation, fatty acid and amino acid metabolism pathways.^{17,23,24} We also want to stress that these severe bronchiolitis endotypes derive from a single cohort and, therefore, warrant further validation, either in another cohort or in functional assays.



Biological pathways from severe bronchiolitis to childhood asthma

Fig. 2. Biological pathways from severe bronchiolitis to childhood asthma. This figure summarizes two biological pathways that may underlie the bronchiolitis endotypes at high risk of developing childhood asthma. From left to right, biological pathway A (i.e., type I/II IFN pathway) links to endotype 3. The type I/II IFN pathway involves the activation of cGAS-STING and the NF-κB-dependent type I/II IFNs and proinflammatory (i.e., IL-6, IL-8) mediators secretion. Additionally, the fatty acid receptor-alveolar macrophage pathway involves the polarization of alveolar macrophages toward an M2 phenotype following activation through FFAR-4. Biological pathway B (i.e., Th2 cell activation and STAT3-glutathione pathways) links to endotype 4. The Th2 cell activation pathway involves an epithelial-derived secretion of pro-Th2 mediators (i.e., IL-33, IL-25, and TSLP) and subsequent differentiation of CD4+ Th0 to Th2 cells. In the presence of ORMDL3 overexpression, the CD4+ Th0 to Th2 cell differentiation is enhanced. In synergy with the Th2 cell activation pathway involves the induction of type I/II IFNs and glutathione derivatives through STAT3- glutathione pathway involves the second messenger cyclic GMP-AMP; cGAMP; cyclic guanosine monophosphate-adenosine monophosphate; FFAR, free fatty acid receptor; IFN, interferon; IL-, interfeukin-; IRF3, interferon regulatory factor 3; IRF7, interferon regulatory of transcription 3; STING, stimulator of interferon genes; Th0, T helper 0; TLR, toll-like receptor; TRIF, toll/interleukin-1R domain-containing adapter-inducing interferon-β; TSLP, thymic stromal lymphopoietin.

Summary

With severe bronchiolitis manifesting as a heterogeneous disease, the quest to define endotypes through the analysis of omics data will help us better assimilate the complexity of this condition. Thus far, bronchiolitis cohort studies have identified distinct severe bronchiolitis endotypes that have differential risks for developing childhood asthma.^{13,16} At this juncture, we believe that there are two representative nasopharyngeal airway endotypes at high risk for childhood asthma development. Specifically, the first representative endotype refers to infants with severe RSV bronchiolitis, a codominance of *M. catarrhalis* and *S. pneumoniae*, upregulated type I/II IFN expression, and altered carbohydrate (i.e., increased mannitol or sorbitol) metabolism. The other endotype refers to infants with severe rhinovirus bronchiolitis, a high abundance of *M. catarrhalis*, upregulated type 2 cytokines, and downregulated sphingolipids metabolism.

With the current data, it remains a challenge to link these endotypes to future precision medicine applications to the management of severe bronchiolitis or to the primary prevention of asthma. However, this scoping review suggests that there are possible approaches to address this challenge. The first approach includes the validation of any identified endotypes in other birth or bronchiolitis cohort studies. The second approach includes the mechanistic validation of endotype-related findings. These approaches will lead to the identification and validation of novel targets for the treatment of severe bronchiolitis, the most common cause of infant hospitalization in the US¹ and a condition still treated with nonspecific interventions.⁷⁴ The proposed work also may provide novel biomarkers for the development of childhood asthma, and thereby facilitate early initiation of asthma treatment for children who develop asthma. More importantly these biomarkers may guide us to novel interventions that would greatly enhance the primary prevention of childhood asthma.

Conflict of interests

The authors state that they have no conflict of interests.

References

- 1. Fujiogi M, Goto T, Yasunaga H, Fujisiro J, Mansbach J, Camargo CA Jr, et al. Trends in bronchiolitis hospitalizations in the United States: 2000–2016. Pediatrics. 2019;144, http://dx.doi.org/10.1542/peds.2019-2614.
- Fujiogi M, Dumas O, Hasegawa K, Jartti T, Camargo CA. Identifying and predicting severe bronchiolitis profiles at high risk for developing asthma: analysis of three prospective cohorts. EClinicalMedicine. 2022;43:101257, http://dx.doi.org/10.1016/j.eclinm.2021.101257.
- Koponen P, Helminen M, Paassilta M, Luukkaala T, Korppi M. Preschool asthma after bronchiolitis in infancy. Eur Respir J. 2012;39:76–80, http://dx.doi.org/10.1183/09031936.00040211.
- Stein RT, Sherrill D, Morgan WJ, Holberg CJ, Halonen M, Taussiget LM, et al. Respiratory syncytial virus in early life and risk of wheeze and allergy by age 13 years. Lancet. 1999;354:541–5, http://dx.doi.org/10.1016/S0140-6736(98)10321-5.
- Dumas O, Erkkola R, Bergroth E, Hasegawa K, Mansbach JM, Piedra PA, et al. Severe bronchiolitis profiles and risk of asthma development in Finnish children. J Allergy Clin Immunol. 2022;149:1281–5, http://dx.doi.org/10.1016/j.jaci.2021.08.035, e1.
- Jackson DJ, Gangnon RE, Evans MD, Roberg KA, Anderson EL, Pappas TE, et al. Wheezing rhinovirus illnesses in early life predict asthma develop-

ment in high-risk children. Am J Respir Crit Care Med. 2008;178:667-72, http://dx.doi.org/10.1164/rccm.200802-309OC.

- Kotaniemi-Syrjanen A, Reijonen TM, Korhonen K, Waris M, Vainionpaa R, Korppi M. Wheezing due to rhinovirus infection in infancy: bronchial hyperresponsiveness at school age. Pediatr Int. 2008;50:506–10, http://dx.doi.org/10.1111/j.1442-200X.2008.02620.x.
- Ralston SL, Lieberthal AS, Meissner HC, Ralston SL, Lieberthal AS, Meissner HC, et al. Clinical practice guideline: the diagnosis, management, and prevention of bronchiolitis. Pediatrics. 2014;134(5):e1474–502, http://dx.doi.org/10.1542/peds.2015-2862.
- 9. Mansbach JM, Qi YS, Espinola JA, Hasegawa K, Puls HT, Sullivan AF, Camargo CA Jr. Late Pre-term Infants with Severe Bronchiolitis and Risk of Asthma by Age 5 Years. J Pediatr. 2022;241:247–50.e1, http://dx.doi.org/10.1016/j.jpeds.2021.09.037.
- Nanishi M, Fujiogi M, Stevenson M, Liang L, Qi YS, Raita Y, et al. Association of Growth Trajectory Profiles with Asthma Development in Infants Hospitalized with Bronchiolitis. J Allergy Clin Immunol Pract. Mar. 2022;10:723–31.e5, http://dx.doi.org/10.1016/j.jaip.2021.11.001.
- Robinson LB, Arroyo AC, Qi YS, Geller RJ, Bauer CS, Hasegawa K, et al. Infant Exposure to Acid Suppressant Medications Increases Risk of Recurrent Wheeze and Asthma in Childhood. J Allergy Clin Immunol Pract. 2022;10:2935–40.e3, http://dx.doi.org/10.1016/j.jaip.2022.07.013.
- Dumas O, Hasegawa K, Mansbach JM, Sullivan AF, Piedra PA, Camargo CA Jr. Severe bronchiolitis profiles and risk of recurrent wheeze by age 3 years. J Allergy Clin Immunol. 2019;143:1371–9.e7, http://dx.doi.org/10.1016/j.jaci.2018.08.043.
- Ooka T, Raita Y, Fujiogi M, Freishtat RJ, Gerszten RE, Mansbach JM, et al. Proteomics endotyping of infants with severe bronchiolitis and risk of childhood asthma. Allergy. 2022;77:3350–61, http://dx.doi.org/10.1111/all.15390.
- Zhu Z, Camargo CA Jr, Raita Y, Freishtat RJ, Fujiogi M, Hahn A, et al. Nasopharyngeal airway dual-transcriptome of infants with severe bronchiolitis and risk of childhood asthma: a multicenter prospective study. J Allergy Clin Immunol. 2022;150:806–16, http://dx.doi.org/10.1016/j.jaci.2022.04.017.
- Altman MC, Kattan M, O'Connor GT, Murphy RC, Whalen E, LeBeau P, et al. Associations between outdoor air pollutants and non-viral asthma exacerbations and airway inflammatory responses in children and adolescents living in urban areas in the USA: a retrospective secondary analysis. Lancet Planet Health. 2023;7:e33–44, http://dx.doi.org/10.1016/S2542-5196(22)00302-3.
- Fujiogi M, Raita Y, Perez-Losada M, Freishtat RJ, Celedón JC, Mansbach JM, et al. Integrated relationship of nasopharyngeal airway host response and microbiome associates with bronchiolitis severity. Nat Commun. 2022;13:4970, http://dx.doi.org/10.1038/s41467-022-32323-y.
- Raita Y, Perez-Losada M, Freishtat RJ, Harmon B, Mansbach JM, Piedra PA, et al. Integrated omics endotyping of infants with respiratory syncytial virus bronchiolitis and risk of childhood asthma. Nat Commun. 2021;12:3601, http://dx.doi.org/10.1038/s41467-021-23859-6.
- Ooka T, Zhu Z, Liang L, Celedon JC, Harmon B, Hahn A, et al. Integrative genetics-metabolomics analysis of infant bronchiolitis-childhood asthma link: a multicenter prospective study. Front Immunol. 2022;13:1111723, http://dx.doi.org/10.3389/fimmu.2022.1111723.
- Shibata R, Zhu Z, Ooka T, Freishtat RJ, Mansbach JM, Pérez-Losada M, et al. Immunoglobulin E-virus phenotypes of infant bronchiolitis and risk of childhood asthma. Front Immunol. 2023;14:1187065, http://dx.doi.org/10.3389/fimmu.2023.1187065.
- Caliskan M, Bochkov YA, Kreiner-Moller E, Bønnelykke K, Stein MM, Du G, et al. Rhinovirus wheezing illness and genetic risk of childhood-onset asthma. N Engl J Med. 2013;368:1398–407, http://dx.doi.org/10.1056/NEJMoa1211592.
- J Med. 2013;368:1398–407, http://dx.doi.org/10.1056/NEJMoa1211592. 21. Kelly RS, Chawes BL, Guo F, Zhang L, Blighe K, Litonjua AA, et al. The role of the 17q21 genotype in the prevention of early childhood asthma and recurrent wheeze by vitamin D. Eur Respir J. 2019;54, http://dx.doi.org/10.1183/13993003.00761-2019.
- Riikonen R, Terasjarvi J, Lauhkonen E, Nuolivirta K, He Q, Korppi M. Interleukin 1 receptor-like 1 rs13408661/13431828 polymorphism is associated with persistent post-bronchiolitis asthma at school age. Acta Paediatr. 2022;111:628–35, http://dx.doi.org/10.1111/apa.16176.
- 23. Raita Y, Camargo CA Jr, Bochkov YA, Celedón JC, Gern JE, Mansbach JM, et al. Integrated-omics endotyping of infants with rhinovirus bronchiolitis and risk of childhood asthma. J Allergy Clin Immunol. 2021;147:2108–17, http://dx.doi.org/10.1016/j.jaci.2020.11.002.
- Zhu Z, Camargo CA Jr, Raita Y, Fujiogi M, Liang L, Rhee EP, et al. Metabolome subtyping of severe bronchiolitis in infancy and risk of childhood asthma. J Allergy Clin Immunol. 2022;149:102–12, http://dx.doi.org/10.1016/j.jaci.2021.05.036.
- Benhamida M, Bihouee T, Verstraete M, Gras Le Guen C, Launay E. Retrospective audit of guidelines for investigation and treatment of bronchiolitis: a French perspective. BMJ Paediatr Open. 2017;1:e000089, http://dx.doi.org/10.1136/bmjpo-2017-000089.
- Kirolos A, Manti S, Blacow R, Tse G, Wilson T, Lister M, et al. A systematic review of clinical practice guidelines for the diagnosis and management of bronchiolitis. J Infect Dis. 2020;222 Suppl. 7:S672–9, http://dx.doi.org/10.1093/infdis/jiz240.
- Dumas O, Mansbach JM, Jartti T, Hasegawa K, Sullivan AF, Piedra PA, et al. A clustering approach to identify severe bronchiolitis profiles in children. Thorax. 2016;71:712–8, http://dx.doi.org/10.1136/thoraxjnl-2016-208535.
- Hasegawa K, Dumas O, Hartert TV, Camargo CA Jr. Advancing our understanding of infant bronchiolitis through phenotyping and endotyping: clinical and molecular approaches. Expert Rev Respir Med. 2016;10:891–9, http://dx.doi.org/10.1080/17476348.2016.1190647.

- Bergroth E, Aakula M, Elenius V, Remes S, Piippo-Savolainen E, Korppi M, et al. Rhinovirus type in severe bronchiolitis and the development of asthma. J Allergy Clin Immunol Pract. 2020;8:588–95, http://dx.doi.org/10.1016/j.jaip.2019.08.043, e4.
- Midulla F, Nicolai A, Ferrara M, Gentile F, Pierangeli A, Bonci E, et al. Recurrent wheezing 36 months after bronchiolitis is associated with rhinovirus infections and blood eosinophilia. Acta Paediatr. 2014;103:1094–9, http://dx.doi.org/10.1111/apa.12720.
- Husby A, Pasanen A, Waage J, Sevelsted A, Hodemaekers H, Janssen R, et al. CDHR3 gene variation and childhood bronchiolitis. J Allergy Clin Immunol. 2017;140:1469–71, http://dx.doi.org/10.1016/j.jaci.2017.06.044, e7.
- Zhou Y, Tong L, Li M, Wang Y, Li L, Yang D, et al. Recurrent wheezing and asthma after respiratory syncytial virus bronchiolitis. Front Pediatr. 2021;9:649003, http://dx.doi.org/10.3389/fped.2021.649003.
- Openshaw PJ, Dean CS, Culley FJ. Links between respiratory syncytial virus bronchiolitis and childhood asthma: clinical and research approaches. Pediatr Infect Dis J. 2003;22 2 Suppl.:S58-64, http://dx.doi.org/10.1097/01.inf.0000053887.26571.eb, discussion S64-5.
- 34. Pala P, Bjarnason R, Sigurbergsson F, Metcalfe C, Sigurs N, Openshaw PJ. Enhanced IL-4 responses in children with a history of respiratory syncytial virus bronchiolitis in infancy. Eur Respir J. 2002;20:376–82, http://dx.doi.org/10.1183/09031936.02.00249902.
- 35. Makrinioti H, Hasegawa K, Lakoumentas J, Xepapadaki P, Tsolia M, Castro-Rodriguez JA, et al. The role of respiratory syncytial virus- and rhinovirus-induced bronchiolitis in recurrent wheeze and asthma – a systematic review and meta-analysis. Pediatr Allergy Immunol. 2022;33:e13741, http://dx.doi.org/10.1111/pai.13741.
- 36. Freid RD, Qi YS, Espinola JA, Cash RE, Aryan Z, Sullivan AF, et al. Proximity to major roads and risks of childhood recurrent wheeze and asthma in a severe bronchiolitis cohort. Int J Environ Res Public Health. 2021;18, http://dx.doi.org/10.3390/ijerph18084197.
- Gong TQ, Jiang YZ, Shao C, Peng WT, Liu MW, Li DQ, et al. Proteome-centric cross-omics characterization and integrated network analyses of triple-negative breast cancer. Cell Rep. 2022;38:110460, http://dx.doi.org/10.1016/j.celrep.2022.110460.
- Yu KH, Snyder M. Omics profiling in precision oncology. Mol Cell Proteomics. 2016;15:2525–36, http://dx.doi.org/10.1074/mcp.0116.059253.
- Zhou Y, Bacharier LB, Isaacson-Schmid M, Baty J, Schechtman KB, Sajolet G, et al. Azithromycin therapy during respiratory syncytial virus bronchiolitis: upper airway microbiome alterations and subsequent recurrent wheeze. J Allergy Clin Immunol. 2016;138:1215–9, http://dx.doi.org/10.1016/j.jaci.2016.03.054, e5.
- Rosas-Salazar C, Shilts MH, Tovchigrechko A, Chappell JD, Larkin EK, Nelson KE, et al. Nasopharyngeal microbiome in respiratory syncytial virus resembles profile associated with increased childhood asthma risk. Am J Respir Crit Care Med. 2016;193:1180–3, http://dx.doi.org/10.1164/rccm.201512-2350LE.
- Mortensen LJ, Kreiner-Moller E, Hakonarson H, Bonnelykke K, Bisgaard H. The PCDH1 gene and asthma in early childhood. Eur Respir J. 2014;43:792–800, http://dx.doi.org/10.1183/09031936.00021613.
- 42. Tutino M, Granell R, Curtin JA, Haider S, Fontanella S, Murray CS, et al. Dog ownership in infancy is protective for persistent wheeze in 17q21 asthma-risk carriers. J Allergy Clin Immunol. 2023;151:423–30, http://dx.doi.org/10.1016/j.jaci.2022.10.012.
- Robinson PFM, Fontanella S, Ananth S, Alonso AM, Cook J, Kaya-de Vries D, et al. Recurrent severe preschool wheeze: from prespecified diagnostic labels to underlying endotypes. Am J Respir Crit Care Med. 2021;204:523–35, http://dx.doi.org/10.1164/rccm.202009-36960C.
- 44. Hoffjan S, Ostrovnaja I, Nicolae D, Newman DL, Nicolae R, Gangnon R, et al. Genetic variation in immunoregulatory pathways and atopic phenotypes in infancy. J Allergy Clin Immunol. 2004;113:511–8, http://dx.doi.org/10.1016/j.jaci.2003.10.044.
- 45. Fujiogi M, Zhu Z, Raita Y, Ooka T, Celedon JC, Freishtat R, et al. Nasopharyngeal lipidomic endotypes of infants with bronchiolitis and risk of childhood asthma: a multicentre prospective study. Thorax. 2022;77:1059–69, http://dx.doi.org/10.1136/thorax-2022-219016.
- Moffatt MF, Kabesch M, Liang L, Dixon AL, Strachan D, Heath S, et al. Genetic variants regulating ORMDL3 expression contribute to the risk of childhood asthma. Nature. 2007;448:470–3, http://dx.doi.org/10.1038/nature06014.
- Wu H, Romieu I, Sienra-Monge JJ, Li H, del Rio-Navarro BE, London SJ. Genetic variation in ORM1-like 3 (ORMDL3) and gasderminlike (GSDML) and childhood asthma. Allergy. 2009;64:629–35, http://dx.doi.org/10.1111/j.1398-9995.2008.01912.x.
- 48. Li X, Howard TD, Zheng SL, Haselkorn T, Peters SP, Meyers DA, et al. Genome-wide association study of asthma identifies RAD50-IL13 and HLA-DR/DQ regions. J Allergy Clin Immunol. 2010;125:328–35, http://dx.doi.org/10.1016/j.jaci.2009.11.018, e11.
- Bonnelykke K, Sleiman P, Nielsen K, Kreiner-Møller E, Mercader JM, Belgrave D, et al. A genome-wide association study identifies CDHR3 as a susceptibility locus for early childhood asthma with severe exacerbations. Nat Genet. 2014;46:51–5, http://dx.doi.org/10.1038/ng.2830.
- Eliasen AU, Pedersen CET, Rasmussen MA, Wang N, Soverini M, Fritz A, et al. Genome-wide study of early and severe childhood asthma identifies interaction between CDHR3 and GSDMB. J Allergy Clin Immunol. 2022;150:622–30, http://dx.doi.org/10.1016/j.jaci.2022.03.019.
- 51. Zhu Z, Hasegawa K, Camargo CA Jr, Liang L. Investigating asthma heterogeneity through shared and distinct genetics: Insights from genome-

wide cross-trait analysis. J Allergy Clin Immunol. 2021;147:796–807, http://dx.doi.org/10.1016/j.jaci.2020.07.004.

- 52. Xu CJ, Soderhall C, Bustamante M, Baïz N, Gruzieva O, Gehring U, et al. DNA methylation in childhood asthma: an epigenome-wide meta-analysis. Lancet Respir Med. 2018;6:379–88, http://dx.doi.org/10.1016/S2213-2600(18)30052-3.
- 53. Arathimos R, Suderman M, Sharp GC, Burrows K, Granell R, Tilling K, et al. Epigenome-wide association study of asthma and wheeze in childhood and adolescence. Clin Epigenet. 2017;9:112, http://dx.doi.org/10.1186/s13148-017-0414-7.
- 54. Reese SE, Xu CJ, den Dekker HT, den Dekker T, Lee H, Sikdar MK, Ruiz-Arenaset S, et al. Epigenome-wide meta-analysis of DNA methylation and childhood asthma. J Allergy Clin Immunol. 2019;143:2062–74, http://dx.doi.org/10.1016/j.jaci.2018.11.043.
- Peters TJ, Buckley MJ, Statham AL, Pidsley R, Samaras K, Lordet RV, et al. De novo identification of differentially methylated regions in the human genome. Epigenet Chromatin. 2015;8:6, http://dx.doi.org/10.1186/1756-8935-8-6.
- Kabesch M, Tost J. Recent findings in the genetics and epigenetics of asthma and allergy. Semin Immunopathol. 2020;42:43–60, http://dx.doi.org/10.1007/s00281-019-00777-w.
- Cardenas A, Sordillo JE, Rifas-Shiman SL, Chung W, Liang L, Coullet BA, et al. The nasal methylome as a biomarker of asthma and airway inflammation in children. Nat Commun. 2019;10:3095, http://dx.doi.org/10.1038/s41467-019-11058-3.
- Donovan BM, Ryckman KK, Breheny PJ, Gebretsadik T, Turi KN, Larkin EK, et al. Association of newborn screening metabolites with risk of wheezing in childhood. Pediatr Res. 2018;84:619–24, http://dx.doi.org/10.1038/s41390-018-0070-4.
- Gurdeniz G, Ernst M, Rago D, Kim M, Courraud J, Stokholm J, et al. Neonatal metabolome of caesarean section and risk of childhood asthma. Eur Respir J. 2022;59, http://dx.doi.org/10.1183/13993003.02406-2021.
- Rago D, Rasmussen MA, Lee-Sarwar KA, Weiss ST, Lasky-Su J, Stokholm J, et al. Fish-oil supplementation in pregnancy, child metabolomics and asthma risk. EBioMedicine. 2019;46:399–410, http://dx.doi.org/10.1016/j.ebiom.2019.07.057.
- Turi KN, McKennan C, Gebretsadik T, Snyder B, Seroogy CM, Lemanske RF Jr, et al. Unconjugated bilirubin is associated with protection from early-life wheeze and childhood asthma. J Allergy Clin Immunol. 2021;148:128–38, http://dx.doi.org/10.1016/j.jaci.2020.12.639.
- Perez-Garcia J, Espuela-Ortiz A, Hernandez-Perez JM, González-Pérez R, Poza-Guedes P, Martin-Gonzalez E, et al. Human genetics influences microbiome composition involved in asthma exacerbations despite inhaled corticosteroid treatment. J Allergy Clin Immunol. 2023, http://dx.doi.org/10.1016/j.jaci.2023.05.021.
- 63. Kozik AJ, Holguin F, Segal LN, Chatila TA, Dixon AE, Gern JE, et al. Microbiome, metabolism, and immunoregulation of asthma: an American Thoracic Society and National Institute of Allergy and Infectious Diseases Workshop Report. Am J Respir Cell Mol Biol. 2022;67:155–63, http://dx.doi.org/10.1165/rcmb.2022-0216ST.
- 64. Poole A, Urbanek C, Eng C, Schageman J, Jacobson J, O'Connor BP, et al. Dissecting childhood asthma with nasal transcriptomics distinguishes subphenotypes of disease. J Allergy Clin Immunol. 2014;133:670–8, http://dx.doi.org/10.1016/j.jaci.2013.11.025, e12.
- 65. Marsh RL, Kaestli M, Chang AB, Binks MJ, Pope CE, Hoffman RE, et al. The microbiota in bronchoalveolar lavage from young children with chronic lung disease

includes taxa present in both the oropharynx and nasopharynx. Microbiome. 2016;4:37, http://dx.doi.org/10.1186/s40168-016-0182-1.

- 66. Thorsen J, Rasmussen MA, Waage J, Waage J, Mortensen M, Brejnrod A, Bønnelykke K, et al. Infant airway microbiota and topical immune perturbations in the origins of childhood asthma. Nat Commun. 2019;10:5001, http://dx.doi.org/10.1038/s41467-019-12989-7.
- Stokholm J, Thorsen J, Blaser MJ, Rasmussen MA, Hjelmsø M, Shah S, et al. Delivery mode and gut microbial changes correlate with an increased risk of childhood asthma. Sci Transl Med. 2020;12, http://dx.doi.org/10.1126/scitranslmed.aax9929.
- Christensen ED, Hjelmso MH, Thorsen J, Shah S, Redgwell T, Poulsen CE, et al. The developing airway and gut microbiota in early life is influenced by age of older siblings. Microbiome. 2022;10:106, http://dx.doi.org/10.1186/s40168-022-01305-z.
- Larkin EK, Gebretsadik T, Moore ML, Anderson LJ, Dupont WD, Chappellet JD, et al. Objectives, design and enrollment results from the Infant Susceptibility to Pulmonary Infections and Asthma Following RSV Exposure Study (INSPIRE). BMC Pulm Med. 2015;15:45, http://dx.doi.org/10.1186/s12890-015-0040-0.
- Kyo M, Zhu Z, Nanishi M, Shibata R, Ooka T, Freishtat RJ, et al. Association of nasopharyngeal and serum glutathione metabolism with bronchiolitis severity and asthma risk: a prospective multicenter cohort study. Metabolites. 2022;12, http://dx.doi.org/10.3390/metabo12080674.
- Raita Y, Perez-Losada M, Freishtat RJ, Hahn A, Castro-Nallar E, Ramos-Tapia I, et al. Nasopharyngeal metatranscriptome profiles of infants with bronchiolitis and risk of childhood asthma: a multicentre prospective study. Eur Respir J. 2022;60, http://dx.doi.org/10.1183/13993003.02293-2021.
- Fujiogi M, Camargo CA Jr, Raita Y, Zhu Z, Celedón JC, Mansbach JM, et al. Integrated associations of nasopharyngeal and serum metabolome with bronchiolitis severity and asthma: a multicenter prospective cohort study. Pediatr Allergy Immunol. 2021;32:905–16, http://dx.doi.org/10.1111/pai.13466.
- McCauley K, Durack J, Valladares R, Fadrosh DW, Lin DL, Calatroni A, et al. Distinct nasal airway bacterial microbiotas differentially relate to exacerbation in pediatric patients with asthma. J Allergy Clin Immunol. 2019;144:1187–97, http://dx.doi.org/10.1016/j.jaci.2019.05.035.
- 74. Pittet LF, Glangetas A, Barazzone-Argiroffo C, Gervaix A, Posfay-Barbe KM, Galetto-Lacouret A, et al. Factors associated with nonadherence to the American Academy of Pediatrics 2014 bronchiolitis guidelines: a retrospective study. PLOS ONE. 2023;18:e0285626, http://dx.doi.org/10.1371/journal.pone.0285626.
- Hallmark B, Wegienka G, Havstad S, Billheimer D, Ownby D, Mendoncaet EA, et al. Chromosome 17q12-21 variants are associated with multiple wheezing phenotypes in childhood. Am J Respir Crit Care Med. 2021;203:864–70, http://dx.doi.org/10.1164/rccm.202003-08200C.
- 76. Zhu Z, Freishtat RJ, Harmon B, Hahn A, Teach SJ, Pérez-Losada M, et al. Nasal airway microRNA profiling of infants with severe bronchiolitis and risk of childhood asthma: a multicentre prospective study. Eur Respir J. 2023;62, http://dx.doi.org/10.1183/13993003.00502-2023.
- 77. Torgerson DG, Giri T, Druley TE, Zheng J, Huntsman S, Seibold MA, et al. Pooled sequencing of candidate genes implicates rare variants in the development of asthma following severe RSV bronchiolitis in infancy. PLOS ONE. 2015;10:e0142649, http://dx.doi.org/10.1371/journal.pone.0142649.
- 78. Törmänen S, Korppi M, Teräsjärvi J, Vuononvirta J, Koponen P, Helminenet M, et al. Polymorphism in the gene encoding toll-like receptor 10 may be associated with asthma after bronchiolitis. Sci Rep. 2017;7:2956, http://dx.doi.org/10.1038/s41598-017-03429-x.